# Seaweed extract and brassinolide can enhance the dwarfing characteristics of potted chrysanthemum

Abou Elhassan, M. H., Bosila, H. A., Hamza, M. A., Elateeq, A. A.\*, Abdel-Gawad, A. I. M. and Soliman, M. N. A.

Horticulture Department, Faculty of Agriculture, Al-Azhar University, Nasr City, Cairo 11754, Egypt.

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Abstract The search for environmental-friendly applications in agriculture is a priority and a necessity. The use of bio-stimulants such as seaweed extracts (SE) and brassinolide (BL) to improve plant growth, quality, and productivity is an effective and eco-friendly strategy. A pot experiment was carried out during the 2018 and 2019 seasons to investigate the effect of spraying plant growth retardants; cycocel (CCC) and paclobutrazol (PPP), in interaction with bio-stimulants; SE (Kelpak®) and BL (Milagrow®) on dwarfing characteristics of chrysanthemum [Dendranthema grandiflorum (Ramat.) Kitamura] cv. 'Pink Zambla' to achieve a desirable plant height with a high dwarfing value. Significant improvements of the studied traits; plant height, leaf number, branche number, plant biomass weight, flowering start, flower number, flower diameter, flowering period, and chlorophyll content was recorded for the combination treatments compared to the single application of the dwarfing factors. Concerning the key characteristics of chrysanthemum as potted plant, the best results were recorded in plants treated by combination of 3000 mg L<sup>-1</sup> CCC or 50 mg L<sup>-1</sup> PPP with different rates of Kelpak or Milagrow. It is possible to recommend the use of growth bio-stimulants along with growth inhibitors to produce a potted dwarf chrysanthemum plant with high flowering quality that increase its commercial value and meets consumer and market needs.

Keywords: Dendranthema, Cycocel, Paclobutrazol, Bio-stimulants, Brassinosteroids

## Introduction

Floriculture is a vital activity in Egypt, and as such, it has made effective economic contributions. In particular, the cultivation of chrysanthemums is of great importance in Egyptian floriculture, and its cultivation has increased throughout the country. The genus *Dendranthema* (DC.) (*Chrysanthemum* L.) belongs to the family Asteraceae and includes about 40 species, widely distributed in China, Mongolia, Japan, and Eastern Europe (Mabberley, 2008;

<sup>\*</sup> Corresponding Author: Elateeq, A. A.; Email: ahmedelateeq@azhar.edu.eg

Youssef et al., 2020). Dendranthema grandiflorum (Ramat.) Kitamura (Chrysanthemum indicum L.) is one of the most important ornamental crops worldwide. Chrysanthemums are commonly used in bouquets and flower arrangements (Vijayakumar et al., 2021). In addition, recently, the importance of this flower as a pot plant has increased in Egypt.

The problem faced by chrysanthemums growers as potted plants is that their height is greater than the desired height beside the irregular growth habit. Controlling plant size is one of the most important aspects of potting plants that can be achieved chemically, genetically, or agronomically (Soliman *et al.*, 2022; Li *et al.*, 2023). Plant growth retardants (PGRs) can be an economical chemical option to control plant height and to improve the quality, balance, and overall appearance of many ornamental and flower plants (Asrar *et al.*, 2014; Toaima *et al.*, 2017; Abou Elhassan *et al.*, 2021). Several reports have shown that there is a wide variation in the sensitivity of chrysanthemum species and cultivars to the application of PGRs (Pobudkiewicz, 2014; Ghatas, 2016; Vaghasia and Polara, 2016).

Among various PGRs, paclobutrazol and cycocel are well known for their use in the production of high-quality potted plants (Soliman et al., 2022). Paclobutrazol (PPP), a triazole plant growth regulator, is effective in controlling vegetative growth and promoting compactness in several ornamental plants (Mishra and Yadava, 2011; Wickramasinghe et al., 2021; Abou Elhassan et al., 2021). Cycocel (CCC) is a synthetic plant growth inhibitor used in ornamental plants to induce dwarfing. In addition to the use of CCC to produce bedding and potted plants, it is used to enhance the green colour of the foliage, strengthen flower stems, and increase the resistance of plants to environmental stresses (Ghatas, 2016). However, damage to plant cells may occur because of cytotoxicity induced by these inhibitors, causing some problems such as delayed flowering, short flowering period, or small flower size. Thus, combining growth stimulants with dwarfing agents may be a good option to mitigate cytotoxicity and overcome this problem. In this context, Hu et al. (2013) found that a mixture application of abscisic acid and the bio-stimulants BL on Leymus chinensis enhanced the net photosynthesis rate, light saturation rate, water use efficiency, stomata conductivity, leaf respiration rate, maximum triphosphate utilization, maximum electron transfer rate, and carboxyl efficiency, thus increased density, plant height, and plant biomass.

Fertilization is one of the most important agricultural practices responsible for increasing the productivity of various crops. However, the improper use of chemical fertilizers by farmers to improve plant growth and productivity is harmful to the environment and human health (Nada *et al.*, 2022). Alternatively, the use of bio-stimulants such as seaweed extracts (SE)

and brassinosteroids (BRs) in agriculture could be a feasible and environmentally friendly approach (Ghatas et al., 2021). Bio-stimulants are a group of substances of natural origin that contribute to the absorption of nutrients, promote plant growth, and increase productivity while reducing dependence on chemical/inorganic fertilizers (Xu and Geelen, 2018). Kelpak, a liquid extract of the seaweed Ecklonia maxima, is a commercial product used as a spray or soil additive for improving plant health, anti-stress, and resistance to pathogenic fungi and nematodes (Oyoo et al., 2010). On the other hand, brassinosteroids are a group of natural steroidal hormones (includes a class of more than 40 polyhydroxylated derivatives) present in plants and play an important role in many biological processes related to growth and development in plants (Kanwar et al., 2022). Milagrow is a commercial natural growth stimulator extracted from the pollen of cabbage flowers (Seadh et al., 2012). Milagrow promotes plant growth, increases yield, improves quality, promotes flower bud formation, and resists flower and fruit drops (Eid et al., 2016). Its composition is 0.2% brassinolide (BL), 20% phosphorous, 10% potassium, and 3% boron (Seadh *et al.*, 2012).

The present study aimed to produce a potted chrysanthemum plant by applying PGRs such as cycocel (CCC) or paclobutrazol (PPP). To improve the dwarfing characteristics and quality of the product enough to meet market quality standards, the growth retardants were combined with growth biostimulants such as seaweed extracts (Kelpak®) and brassinolide (Milagrow®) as environmentally friendly natural stimulants instead of chemical fertilization. The vegetative growth, flowering aspects, and photosynthetic pigments of *D. grandiflorum* cv. 'Pink Zambla' were considered in the current study.

#### Materials and methods

The experiment was conducted in a private farm at El-Qurin (30 °35'17.1"N 31 °44'45.8"E), Sharkia, Egypt, during the two successive seasons 2018 and 2019. The experiment aimed to investigate the effect of the PGRs; CCC and PPP alone or in combination with the growth bio-stimulants; seaweed extracts (SE) (Kelpak®) and brassinolide (BL) (Milagrow®) on the dwarfing characteristics of *D. grandiflorum* (Ramat.) Kitamura cv. 'Pink Zambla'.

#### Plant materials

*D. grandiflorum* cv. 'Pink Zambla' cuttings were obtained from a private farm at El-Qanater El-Khayreya, Qalyubia, Egypt. Terminal cuttings (8-10 cm long) were treated with indole 3-butyric acid (IBA) at 2000 mg L<sup>-1</sup>. The

cuttings were then planted in a mixed medium of sand and peat moss in a ratio of 1:1 (v/v) and were incubated under long-day conditions and relatively high humidity (mist) for 10-15 days until fully rooting.

## Mixture substrate (soil)

A commercial mixture substrate composed of 20% perlite and 80% peat moss (v/v) was prepared homogeneity before cultivation. The substrate pH was adjusted to 6.2 with calcium carbonate. The mixture substrate was chemically analyzed at National Research Centre, Dokki, Cairo, Egypt, and the results are presented in Table (1).

**Table 1.** Physical and chemical properties of the experimental mixture substrate (soil)

Items	1 <sup>st</sup> season	2 <sup>nd</sup> season	
Moisture %	38	43	
Ash %	51.61	63	
Organic matter %	10.39	9.00	
pН	7	6.3	
E.C ds.m <sup>-1</sup>	2.35	2.38	
CaCO <sub>3</sub> %	2.85	2.60	
A	vailable macronutrients (mg kg <sup>-1</sup> so	oil)	
N	50	51	
P	11	13	
K	78	80	
Ca	1200	1142	
Mg	162	170	
Na	144	139	
A	vailable micronutrients (mg kg <sup>-1</sup> so	oil)	
Fe	4.3	4.1	
Mn	4.4	4	
Zn	4.5	4.1	
Cu	0.8	0.6	

## Cultivation and agricultural practices

Pots of 14 cm diameter were filled with the previously prepared mixture. The pots were arranged into groups of 15 pots for each treatment which contains 3 replicates of 5 pots each. The different groups of the prepared pots were placed in the spacing of 30 cm x 30 cm. Uniform rooted cuttings (15 days old) were planted on February 1<sup>st</sup> in both seasons. Containers containing plants (one plant per pot) were grown under a Multi Span greenhouse covered with plastic (120 microns thickness). Chrysanthemums are short-day plants as they

only bloom in short daylight conditions. Therefore, flowering time can be regulated by controlling the photocycle to give marketable flowers during the year. Artificial light for 4 h (3 meters between lamps and 1.75 meters between plant and lamp) was employed to prolong the day for more than 16 h for all treatments for 28 days. After that, plants were covered with black plastic from 5 PM to 7 AM until color appeared in the flower bud. Pinching was carried out 2 weeks after transplanting. Plants were watered by drip irrigation to control irrigation management with a frequency depending on weather and plant conditions.

#### **Treatments**

## The growth stimulants

SE (Kelpak<sup>®</sup>) was sprayed at 0, 3, 4, and 5 ml L<sup>-1</sup>, while BL (Milagrow<sup>®</sup>) was applied at 0, 60, 80, and 100 mg L<sup>-1</sup>. Both stimulants were sprayed 4 times at 3, 5, 7, and 9 weeks from transplanting. The composition of Kelpak<sup>®</sup> is displayed in Table (2) according to Anton *et al.* (2010). Milagrow is a registered trademark product of Green India Co., India. It has been obtained from Jaara Company, Cairo, Egypt. The composition of Milagrow is 0.2% BL, 20% phosphorous, 10% potassium, and 3% boron (Seadh *et al.*, 2012).

**Table 2.** Chemical composition of Kelpak®

Composition (mg L <sup>-1</sup> )										
Auxins	10.7	Cytokinins	0.03	Amino acids	1000	Carbohydrates	10000			
Proteins	2000	$P_2O_5$	300	Mg	56.4	Ca	200			
Tot N	400	$K_2O$	6100	Mn	0.8	В	3.2			
Org N	4000	Fe	2.2	Cu	1.8	Zn	0.9			

#### The growth retardants

CCC (cycocel, or 2-chloro ethyl trimethyl ammonium chloride) at rates of 1500, 3000, and 6000 mg L<sup>-1</sup>, and PPP (paclobutrazol) at 25, 50, and 75 mg L<sup>-1</sup> were applied 4 weeks after transplanting (2 weeks after pinching) and repeated 2 weeks later. The spray solution soaked both leaves and stems till running off point using a 2-liter hand pump sprayer. The treatments were applied in the afternoon (4.00 pm) for easier absorption.

#### Measurements

Plant height (cm) from the soil surface to stem apex, leaf number per plant, and branche number per plant were recorded at the beginning of flowering. Fresh weight (g plant<sup>-1</sup>) of aerial parts including flowers was

recorded at the stage of complete flowering opening. Dry weight (g plant<sup>-1</sup>) was determined after drying in an oven at 65°C for 48 h.

Flowering start (days) was considered at the first bud sprouting. The flower number per plant was calculated for both opened and unopened flowers. Flower diameter (cm) was measured for fully opened flowers. The flowering period (days) was recorded from the first opened flower until the end of flowering.

The content of chlorophyll and carotenoids (mg g<sup>-1</sup> FW) were determined according to Lichtenthaler (1987). About 0.2 g of the fresh leaf was mixed with 15 ml acetone (80%). After filtration, the volume was adjusted to 15 ml with acetone (80%), and the absorption was measured with a JENWAY 6800 UV/Vis. spectrophotometer at 663.2, 646.8, and 470 nm against acetone (80%) blank. The concentration of chlorophyll (Chl) and carotenoids (Car) was calculated using the following formula:

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\begin{split} & Chl.a = 12.25A_{663.2} - 2.79A_{646.8} \\ & Chl.b = 21.50A_{646.8} - 5.1A_{663.2} \\ & Total \ Chl = 7.15A_{663.2} + 18.71A_{646.8} \\ & Car = (1000A_{470} - 1.8Chl.a - 85.02Chl.b) \ / \ 198 \end{split}
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## Statistical analysis

The experiment design was arranged in a randomized complete design during the two seasons. Each treatment contained three replicates and each replicate consisted of 5 potted plants. The statistical analysis of data was subjected to Analysis of Variance (ANOVA), and means were compared by L.S.D. at 5% using COSTAT package ver. 6.4 (CoHort software Monterey, USA) according to Snedecor and Cochran (1980).

#### Results

## Plant height (cm)

Spraying the dwarfing agents, CCC and PPP, on chrysanthemum plants resulted in a significant reduction in plant height compared to control treatment in both seasons (Figure 1A). Plant height was decreased linearly with the increase in the concentration of CCC and PPP. Higher dosages of CCC (6000 mg L<sup>-1</sup>) and PPP (75 mg L<sup>-1</sup>) severely reduced the plant height to 12.00 and 14.67 cm in the 2018 season, and to 14.00 and 14.67 cm in the 2019 season, respectively. Moderate applications of dwarfing agents (CCC 3000 mg L<sup>-1</sup> and PPP 50 mg L<sup>-1</sup>) achieved stem lengths (20-25 cm) close to commercially desirable plant lengths (around 30 cm).

Within the combination treatments of plant retardants; CCC and PPP, and growth stimulants; Kelpak and Milagrow, the highest significant values of plant height (59.45, 60.53, 59.40, 59.43, and 61.60 cm) were recorded when 3, 4, and 5 ml L<sup>-1</sup> Kelpak, and 80 and 100 mg L<sup>-1</sup> Milagrow were sprayed to plants dwarfed with 25 mg L<sup>-1</sup> PPP in the 1<sup>st</sup> season, respectively, without significant differences between them (Table 3). In the 2<sup>nd</sup> season, the same trend was observed. The suitable commercial length for chrysanthemum plants (25-35 cm) was gained when 3000 mg L<sup>-1</sup> of CCC or 50 mg L<sup>-1</sup> of PPP was applied in interaction with the growth stimulants Kelpak and Milagrow at any rate.

**Table 3.** Effect of cycocel (CCC) and paclobutrazol (PPP) in interaction with Kelpak and Milagrow on plant height of chrysanthemum

Kelpak and Winagrow on plant height of em ysanthemum											
Plant height (cm)											
First season (2018)											
Treatm	onte	Control	Ke	lpak (ml	$L^{-1}$ )	Mila	Milagrow (mg L <sup>-1</sup> )				
1164111	ents	0	3	4	5	60	80	100	Mean A		
CCC	1500	49.33	49.97	51.23	52.91	48.83	53.13	55.53	51.56		
CCC	3000	20.00	27.13	29.13	30.23	28.13	29.27	32.18	28.01		
$(\text{mg L}^{-1})$	6000	12.00	12.67	14.13	14.63	13.50	15.72	16.38	14.15		
PPP	25	55.00	59.45	60.53	59.40	57.32	59.43	61.60	58.96		
	50	23.01	30.85	32.00	32.48	30.20	31.23	34.23	30.57		
$(\text{mg L}^{-1})$	75	14.67	15.52	15.03	16.92	16.05	17.08	18.72	16.28		
Mean B		29.00	32.60	33.68	34.43	32.34	34.31	36.44			
L.S.D. at 5	%: A = 0	.94, B=1.02	2, A*B=2	.49							
			Se	cond seas	son (2019	))					
CCC	1500	50.33	50.43	53.33	53.40	49.87	52.83	57.80	52.57		
	3000	22.00	27.60	30.50	29.67	30.87	30.07	34.00	29.24		
$(\text{mg L}^{-1})$	6000	14.00	16.59	17.33	16.88	14.96	16.44	17.96	16.31		
PPP	25	70.00	74.85	75.29	76.00	75.33	78.48	79.40	75.62		
	50	25.00	30.11	33.59	34.18	28.67	33.11	37.22	31.70		
$(\text{mg L}^{-1})$	75	14.67	15.81	16.07	17.92	16.22	17.51	21.29	17.07		
Mean B		32.67	35.90	37.69	38.01	35.99	38.07	41.28			
L.S.D. at 5	%: A= 1	.23, B=1.33	8, A*B=3	.26							

## Leaf number per plant

As displayed in Figure (1B), treating chrysanthemum plants with CCC and PPP reduced leaf number compared with the control. The negative effect of CCC and PPP on leaves number was increased with the increase in dwarfing agent level. The lowest leaf number (9.33 and 11.00 leaves per plant) were counted for plants dwarfed by the higher dose of PPP (75 mg  $L^{-1}$ ) in the first and second seasons, respectively, followed by a higher level of CCC (6000 mg  $L^{-1}$ ).

A significant improvement was observed in the leaf number of dwarfed plants when they were sprayed with Kelpak or Milagrow (Table 4). The highest significant value of leaf number in 1<sup>st</sup> season was recorded for the treatment of CCC (1500 mg L<sup>-1</sup>) with Milagrow (100 mg L<sup>-1</sup>) (63.33 leaves per plant) and the treatment of PPP (25 mg L<sup>-1</sup>) with Milagrow (100 mg L<sup>-1</sup>) which recorded 58.00 leaves per plant.

**Table 4.** Effect of cycocel (CCC) and paclobutrazol (PPP) in interaction with Kelpak and Milagrow on leaf number per plant of chrysanthemum

Telpak und Muggew en rear hander per plant er ein yearthemen												
	Leaf number per plant											
					on (2018							
Treatm	onte	Control	Kelpak (ml L		$L^{-1}$	Milagrow (mg L <sup>-1</sup> )			Mean A			
Heatin	CIIIS	0	3	4	5	60	80	100				
CCC	1500	42.00	46.67	53.00	56.67	56.00	57.67	63.33	53.62			
	3000	35.33	37.00	38.00	45.00	49.33	52.33	53.33	44.33			
$(\text{mg L}^{-1})$	6000	12.00	17.67	19.00	25.00	21.00	22.00	22.67	19.90			
DDD	25	42.00	49.33	51.00	52.67	54.00	54.67	58.00	51.67			
PPP	50	28.33	35.33	37.67	39.67	37.67	41.33	42.33	37.48			
$(\text{mg L}^{-1})$	75	9.33	23.33	26.67	27.67	28.67	30.67	33.67	25.71			
Mean B		28.17	34.89	37.56	41.11	41.11	43.11	45.56				
L.S.D. at 5	5%: A= 2	2.08, B = 2.2	4, A*B=	5.49								
			S	econd sea	ason (201	9)						
CCC	1500	45.00	52.33	57.00	60.67	64.33	66.00	74.33	59.95			
CCC	3000	34.33	40.67	43.67	48.67	51.33	55.00	59.33	47.57			
$(\text{mg L}^{-1})$	6000	11.33	20.67	24.67	29.33	25.00	26.33	27.67	23.57			
DDD	25	44.00	48.67	54.67	58.33	59.33	63.00	66.00	56.29			
PPP	50	26.00	37.67	39.33	40.67	39.00	42.33	52.67	39.67			
$(\text{mg L}^{-1})$	75	11.00	22.33	29.67	32.33	31.67	34.67	39.67	28.76			
Mean B		28.61	37.06	41.50	45.00	45.11	47.89	53.28				
L.S.D. at 5	L.S.D. at 5%: A= 2.43, B= 2.63, A*B= 6.44											

## Branche number per plant

Application of CCC at low (1500 mg L<sup>-1</sup>) and moderate (3000 mg L<sup>-1</sup>) concentrations and PPP at a moderate level (50 mg L<sup>-1</sup>) improved branching as compared to untreated plants (Figure 1C). The highest number of branches (4.00 branches per plant) was produced on plants treated with 1500 mg L<sup>-1</sup> CCC in the 2018 season followed by 3000 mg L<sup>-1</sup> CCC and 50 mg L<sup>-1</sup> PPP (3.67 branches per plant). The same treatments recorded the highest branching in the 2019 season. Higher dosages of the dwarfing agents resulted in sever inhibition in branching.

An improvement in chrysanthemum branching was observed in the combination treatment (Table 5). Herein, in the second season, the highest values of branche number (4.40, 4.67, and 4.46 branches per plant) were

noticed when 100 mg  $L^{-1}$  of Milagrow interacted with 1500, 3000 mg  $L^{-1}$  CCC, and 50 mg  $L^{-1}$  PPP, respectively.

## Fresh and dry weight (g plant 1)

Fresh (FW) and dry weight (DW) were decreased in plants treated with high level of CCC or PPP separately (Figure 1D and 1E). The dwarfing caused by low and medium concentrations of CCC did not reduce the FW significantly when compared to control. In contrast, treating plants with 25 and 50 mg L<sup>-1</sup> of PPP increased FW but insignificantly compared to control in both seasons. Low concentration of PPP (25 mg L<sup>-1</sup>) recorded the highest significant DW in the first season (11.73 versus 11.17 g plant<sup>-1</sup> of control).

**Table 5.** Effect of cycocel (CCC) and paclobutrazol (PPP) in interaction with Kelpak and Milagrow on branche number per plant of chrysanthemum

Branche number per plant First season (2018)											
Treatm	ents	Control		elpak (ml	. /		agrow (m	g L <sup>-1</sup> )	Mean A		
		0	3	4	5	60	80	100	_		
CCC	1500	4.00	4.00	4.03	4.00	4.33	3.87	4.47	4.10		
CCC	3000	3.67	3.70	3.93	4.33	3.83	4.33	4.33	4.02		
$(\text{mg L}^{-1})$	6000	1.33	1.70	1.77	1.87	1.40	1.67	1.50	1.60		
PPP	25	2.67	3.23	3.33	3.43	3.56	3.60	3.33	3.31		
	50	3.67	4.30	4.33	4.57	3.50	3.90	4.40	4.10		
$(\text{mg L}^{-1})$	75	1.67	2.00	2.33	2.43	1.63	1.60	2.33	2.00		
Mean B		2.83	3.16	3.29	3.44	3.04	3.16	3.39			
L.S.D. at 59	%: A= 0.	14, B = 0.15	5, A*B=0	0.37							
			Se	cond sea	son (2019	9)					
CCC	1500	3.67	3.43	3.67	4.33	4.33	4.17	4.40	4.00		
	3000	3.67	3.67	4.00	4.23	4.00	4.10	4.67	4.05		
$(\text{mg L}^{-1})$	6000	1.27	2.00	2.00	1.90	2.00	2.33	1.77	1.90		
PPP	25	2.33	3.07	3.23	3.63	3.47	3.43	3.00	3.17		
	50	3.67	4.13	4.17	4.20	3.67	4.33	4.46	4.09		
$(\text{mg L}^{-1})$	75	1.77	1.83	2.07	1.93	1.80	1.90	2.20	1.93		
Mean	В	2.73	3.02	3.19	3.37	3.21	3.38	3.42			
L.S.D. at 59	%: A= 0.	10, B = 0.10	), A*B= (	0.25							

Regarding the interaction treatments, data tabulated in Table (6 and 7) revealed that the FW and DW of plants treated with Kelpak and Milagrow and dwarfed with CCC and PPP was enhanced in most treatments.

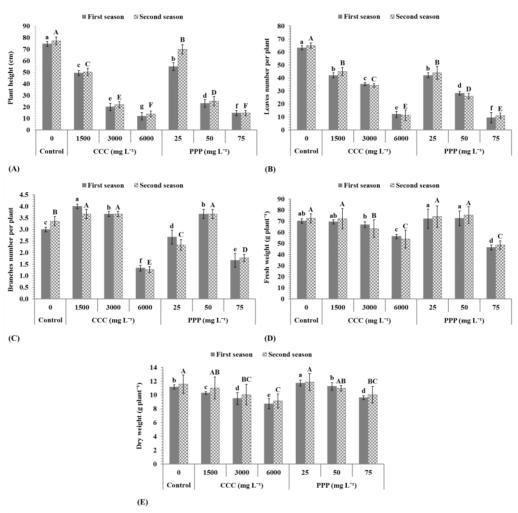
**Table 6.** Effect of cycocel (CCC) and paclobutrazol (PPP) in interaction with Kelpak and Milagrow on fresh weigh of chrysanthemum

Kelpak and Milagrow on Iresh weigh of chrysanthemum											
	Fresh weight (g plant <sup>-1</sup> )										
First season (2018)											
Tractma	onto	Control	K	elpak (ml	L-1)	Mila	Milagrow (mg L <sup>-1</sup> )				
Treatm	ents	0	3	4	5	60	80	100	Mean A		
CCC	1500	69.50	71.89	77.03	85.00	78.49	84.78	87.93	79.23		
$CCC$ $(mg L^{-1})$	3000	67.00	68.30	68.73	73.59	71.82	74.85	81.87	72.31		
(IIIg L )	6000	56.33	58.28	66.05	70.79	60.12	61.77	64.92	62.61		
PPP	25	72.30	77.52	78.79	83.17	76.82	78.10	80.96	78.23		
	50	72.72	75.02	76.91	79.37	72.89	75.48	83.74	76.59		
$(\text{mg L}^{-1})$	75	46.33	58.09	61.38	64.71	52.36	57.86	64.08	57.83		
Mean B		64.03	68.19	71.48	76.11	68.75	72.14	77.25			
L.S.D. at 5	5%: A= 2	2.19, B=2.	37, A*B=	5.80							
			S	Second sea	ason (2019	9)			_		
CCC	1500	72.28	74.80	79.95	87.52	77.60	89.45	96.37	82.57		
CCC	3000	63.42	67.59	68.46	72.69	70.93	78.43	83.79	72.19		
$(\text{mg L}^{-1})$	6000	54.00	55.94	65.40	68.04	58.96	63.32	66.67	61.76		
PPP	25	74.30	79.16	81.82	92.26	78.83	81.99	83.89	81.75		
	50	75.58	79.26	79.70	77.42	69.80	76.59	75.45	76.26		
$(\text{mg L}^{-1})$	75	48.55	63.45	66.59	65.41	59.23	62.98	65.08	61.61		
Mean	В	64.69	70.03	73.66	77.22	69.23	75.46	78.54			
L.S.D. at 5	5%: A= 2	2.42, B=2.	62, A*B=	6.42							

**Table 7.** Effect of cycocel (CCC) and paclobutrazol (PPP) in interaction with Kelpak and Milagrow on dry weight of chrysanthemum

$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	Dry weight (g plant <sup>-1</sup> )										
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$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	A										
$\begin{array}{cccccccccccccccccccccccccccccccccccc$											
PPP 25 11.73 12.15 12.21 12.95 11.99 12.19 12.62 12.26 (mg I -1) 50 11.26 11.32 11.63 11.88 10.30 10.95 12.21 11.36											
$\frac{PPP}{(mg I^{-1})}$ 50 11.26 11.32 11.63 11.88 10.30 10.95 12.21 11.36											
$(\text{mg I}^{-1})$ 50 11.26 11.32 11.63 11.88 10.30 10.95 12.21 11.36											
(ling L ) 75 9.61 11.18 11.83 12.61 10.53 10.75 10.93 11.06											
Mean B 10.19 10.64 11.03 11.74 10.39 10.80 11.35											
L.S.D. at 5%: A= 0.35, B= 0.38, A*B= 0.93											
Second season (2019)											
CCC 1500 11.03 10.67 11.49 11.85 11.67 12.41 13.31 11.77											
$(\text{mg L}^{-1})$ 3000 10.06 10.54 10.75 12.22 9.72 10.06 12.01 10.77											
(ling L ) 6000 9.14 8.83 9.47 10.03 7.78 9.00 9.35 9.09											
PPP 25 11.89 12.64 12.56 13.09 12.23 12.81 13.26 12.64											
$(\text{mg L}^{-1})$ 50 10.99 11.37 12.09 12.10 9.71 10.53 11.14 11.13											
(lig L ) 75 10.05 11.46 12.19 12.17 10.88 10.83 11.12 11.24											
Mean B 10.53 10.92 11.43 11.91 10.33 10.94 11.70											
L.S.D. at 5%: A= 0.40, B= 0.43, A*B= 1.05											

Plant FW reached the highest value when the combination of 1500 mg  $L^{-1}$  CCC and 100 mg  $L^{-1}$  Milagrow was applied, which achieved 87.93 and 96.37 g plant<sup>-1</sup> in the first and second seasons, respectively. The highest DWs were found for 1500 mg  $L^{-1}$  CCC with 100 mg  $L^{-1}$  Milagrow and 25 mg  $L^{-1}$  PPP with 5 ml  $L^{-1}$  Kelpak.



**Figure 1.** Effect of different concentrations of cycocel (CCC) and paclobutrazol (PPP) on vegetative characters: plant height (cm) (A), leaf number per plant (B), branche number per plant (C), fresh weight (g plant<sup>-1</sup>) (D), and dry weight (g plant<sup>-1</sup>) (E) of *D. grandiflorum* cv. 'Pink Zambla' during the seasons of 2018 and 2019. Bars represent ±Standard Deviation (n=3). Columns annotated with the same letters are not statistically different (*P*<0.05)

## Flowering start (days)

CCC and PPP applied separately caused significantly earlier flowering compared to the control in both seasons except for treatment 75 mg L<sup>-1</sup> PPP, which delayed flowering as compared with the control by 5 days and 1 day in the 1<sup>st</sup> and 2<sup>nd</sup> seasons, respectively (Figure 2A). In this context, the earlier flowering was achieved due to applying medium concentrations of CCC (3000 mg L<sup>-1</sup>) and PPP (50 mg L<sup>-1</sup>) (85.33 and 82.33 days in the 1<sup>st</sup> season, and 82.33 and 84.33 days in the 2<sup>nd</sup> season, respectively).

The interaction between PGRs; CCC and PPP, and the growth stimulants; Kelpak and Milagrow showed a positive effect on the time of the flowering start of the chrysanthemum plant (Table 8). The earliest flowering was achieved when 3000 mg L<sup>-1</sup> of CCC was combined with 100 mg L<sup>-1</sup> of Milagrow, as the plants flowered after 80.27 and 80.07 days of cultivation, in the 2018 and 2019 seasons, respectively. This means that these plants can be produced 2-5 days earlier than plants stunted with 3000 mg L<sup>-1</sup> CCC only (85.33 and 82.33 days in the first and second seasons, respectively).

**Table 8.** Effect of cycocel (CCC) and paclobutrazol (PPP) in interaction with Kelpak and Milagrow on flowering start of chrysanthemum

Flowering start (days) First season (2018)										
		Control	K	elpak (ml			agrow (mg	g L <sup>-1</sup> )	3.6	
Treatme	ents	0	3	4	5	60	80	100	- Mean A	
CCC	1500	98.33	87.13	86.00	84.80	87.26	87.67	85.87	88.15	
$(\text{mg L}^{-1})$	3000	85.33	84.73	84.33	83.87	83.47	82.27	80.27	83.47	
(IIIg L )	6000	95.33	89.93	88.00	85.13	92.13	87.33	85.67	89.08	
PPP	25	92.67	90.00	88.53	87.13	84.93	82.27	82.07	86.80	
$(\text{mg L}^{-1})$	50	82.33	82.33	81.80	82.27	83.33	83.83	81.13	82.43	
(mg L )	75	105.33	92.13	89.67	89.20	87.27	86.20	85.13	90.70	
Mean	В	93.22	87.71	86.39	85.40	86.40	84.93	83.36		
L.S.D. at 5	5%: A= (	0.50, B = 0.	54, A*B=	: 1.33						
			5	Second sea	ason (2019	9)				
CCC	1500	88.67	86.47	85.13	84.07	85.47	84.47	82.07	85.19	
$(\text{mg L}^{-1})$	3000	82.33	82.73	82.40	81.27	81.20	80.60	80.07	81.52	
(mg L )	6000	92.00	88.80	87.27	85.20	91.20	86.60	86.00	88.15	
PPP	25	90.00	88.13	86.80	86.07	86.27	84.13	82.13	86.22	
$(\text{mg L}^{-1})$	50	84.33	81.87	81.93	81.80	83.73	82.60	80.60	82.41	
(mg L )	75	102.33	91.73	89.80	88.53	88.67	87.07	85.20	90.48	
Mean	В	89.94	86.62	85.56	84.49	86.10	84.24	82.68		
L.S.D. at 5	5%: A= (	0.55, B = 0.	59, A*B=	1.45						

## Flower number per plant

Application of CCC and PPP each alone at a moderate dose improved flower number as compared to non-dwarfed plants. The highest significant number of flowers was produced from plants treated with 3000 mg L<sup>-1</sup> of CCC (12.10 and 12.90 flowers per plant) and 50 mg L<sup>-1</sup> of PPP (10.93 and 11.93 flowers per plant) in 2018 and 2019 seasons, respectively, compared to control (9.47 and 10.03 flowers per plant) (Figure 2B).

Regarding the interaction treatment, the highest significant number of flowers (14.29 and 16.07 flowers per plant) was counted for the treatment of 3000 mg L<sup>-1</sup> of CCC in interaction with Milagrow at 100 mg L<sup>-1</sup> compared to the single spraying with 3000 mg L<sup>-1</sup> CCC (12.10 and 12.90 flowers per plant) in the first and second seasons, respectively (Table 9). The following combinations also gained the highest production of flowers without significant differences between them: 1500 mg L<sup>-1</sup> CCC with 100 mg L<sup>-1</sup> Milagrow, 3000 mg L<sup>-1</sup> CCC with 4 and 5 ml L<sup>-1</sup> Kelpak, 50 mg L<sup>-1</sup> PPP with 5 ml L<sup>-1</sup> Kelpak and 100 mg L<sup>-1</sup> Milagrow, in the first season. Within different combinations, flower number was increased by the raising rate of Kelpak and Milagrow.

**Table 9.** Effect of cycocel (CCC) and paclobutrazol (PPP) in interaction with Kelpak and Milagrow on flower number of chrysanthemum

Respar and whagrow on nower number of chrysanthemum												
	Flower number per plant First season (2018)											
Tuanton	-mta	Control	l Kelpak (ml L <sup>-1</sup> )		Mila	Milagrow (mg L <sup>-1</sup> )						
Treatme	ents	0	3	4	5	60	80	100	-			
CCC	1500	10.10	12.52	12.92	12.96	13.17	13.30	13.95	12.70			
CCC	3000	12.10	12.78	13.71	14.13	13.32	13.64	14.29	13.42			
$(\text{mg L}^{-1})$	6000	2.97	3.28	3.35	3.62	3.13	3.58	4.00	3.42			
PPP	25	8.00	9.68	9.66	10.27	10.77	10.98	11.00	10.05			
	50	10.93	12.08	12.71	14.17	12.65	13.35	14.16	12.87			
$(\text{mg L}^{-1})$	75	2.57	3.68	3.98	4.32	3.78	3.85	4.15	3.76			
Mean	В	7.78	9.00	9.39	9.91	9.47	9.78	10.26				
L.S.D. at 5	%: A = 0	0.24, B = 0.2	26, A*B=	0.63								
			S	Second sea	ason (2019	9)						
CCC	1500	9.10	12.88	13.58	13.80	13.32	13.73	14.25	12.95			
(mg L <sup>-1</sup> )	3000	12.90	13.88	14.10	15.12	14.02	14.78	16.07	14.41			
(mg L )	6000	2.10	3.55	3.70	3.29	3.23	3.62	3.75	3.32			
PPP	25	8.90	10.00	9.98	11.02	10.95	11.18	11.65	10.53			
$(\text{mg L}^{-1})$	50	11.93	12.95	13.37	14.30	13.00	14.03	15.10	13.52			
(IIIg L )	75	2.10	3.49	3.78	4.37	3.95	4.27	4.50	3.78			
Mean	В	7.84	9.46	9.75	10.32	9.74	10.27	10.89				
L.S.D. at 5	%: A= 0	0.26, B = 0.2	28, A*B=	0.69								

## Flower diameter (cm)

The data presented in Figure (2C) revealed that the chrysanthemum plants dwarfed by CCC and PPP each alone produced flowers smaller than those of non-dwarfed plants in both seasons. A slight increase in flower diameter (4.47 and 4.57 cm) was noticed for CCC application at 3000 mg L<sup>-1</sup> in the 1<sup>st</sup> and 2<sup>nd</sup> seasons, respectively, without significant differences with the control (4.40 and 4.43 cm).

The flower size was increased upon spraying the bio-stimulants (Table 10). In the first season, the largest sizes of flower (5.47, 5.87, 5.77, and 6.03 cm) were recorded when 4 ml L<sup>-1</sup> Kelpak, 80 or 100 mg L<sup>-1</sup> of Milagrow were sprayed on plants dwarfed with 3000 mg L<sup>-1</sup> of CCC, as well as when 100 mg L<sup>-1</sup> of Milagrow applied on plants stunted with 50 mg L<sup>-1</sup> of PPP, respectively, without significant differences between them. In the second season, the following combinations also resulted in the highest value for flower diameter without statistical differences: 3000 mg L<sup>-1</sup> CCC with 4 or 5 ml L<sup>-1</sup> Kelpak, 80 or 100 mg L<sup>-1</sup> Milagrow; 25 mg L<sup>-1</sup> PPP with 5 ml L<sup>-1</sup> Kelpak or 100 mg L<sup>-1</sup> Milagrow.

## Flowering period (days)

There was no significant increase in the flowering period of dwarfed chrysanthemum plants in the first season (Figure 2D). In the second season, only 3000 mg L<sup>-1</sup> CCC recorded a significant increase in the flowering period when compared with non-dwarfed plants (42.23 versus 35.93 days). The longest periods of flowering (44.20 and 42.23 days) were observed for 3000 mg L<sup>-1</sup> of CCC in the first and second seasons, respectively.

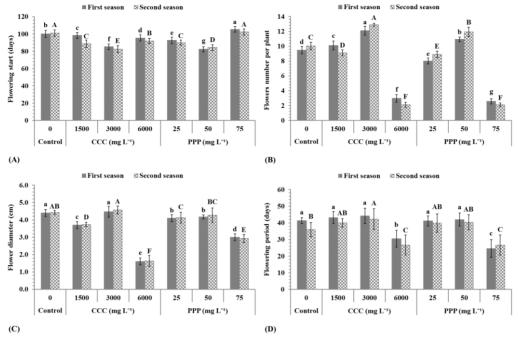
The treatment of moderate concentrations of CCC (3000 mg L<sup>-1</sup>) and PPP (50 mg L<sup>-1</sup>) combined with different rates of Kelpak and Milagrow achieved a significant prolongation of the flowering period (Table 11). Herein, the flowering period was increased up to 11 days than plants received the dwarfing agents only. In the 1<sup>st</sup> season, the longest significant periods of flowering were recorded when 100 mg L<sup>-1</sup> of Milagrow was applied to plants treated with 1500 mg L<sup>-1</sup> CCC (49.67 days), 3000 mg L<sup>-1</sup> CCC (52.20 days), 25 mg L<sup>-1</sup> PPP (48.83 days), or 50 mg L<sup>-1</sup> PPP (50.00 days), beside combination of 5 ml L<sup>-1</sup> Kelpak with 3000 mg L<sup>-1</sup> CCC (50.50 days) or with 50 mg L<sup>-1</sup> PPP (48.63 days), without significant differences between them. The same trend of combinations was observed in the second season.

**Table 10.** Effect of cycocel (CCC) and paclobutrazol (PPP) in interaction with Kelpak and Milagrow on flower diameter of chrysanthemum

Kelpak and whiagrow on hower diameter of emysanthemum											
Flower diameter (cm)											
First season (2018)											
Treatme	onto	Control	Kelpak (ml L <sup>-1</sup> )			Mila	Milagrow (mg L <sup>-1</sup> )				
Treating	ents	0	3	4	5	60	80	100	_		
CCC	1500	3.70	3.90	4.07	4.27	4.37	4.43	5.00	4.25		
CCC	3000	4.47	5.23	5.47	5.30	5.10	5.87	5.77	5.31		
$(\text{mg L}^{-1})$	6000	1.60	2.50	2.97	3.03	2.30	2.37	3.00	2.54		
PPP	25	4.10	4.67	5.10	5.27	4.77	4.73	5.00	4.80		
$(\text{mg L}^{-1})$	50	4.17	4.60	4.63	4.80	4.87	5.30	6.03	4.91		
(IIIg L )	75	3.00	3.10	3.33	3.30	3.10	3.17	3.23	3.18		
Mean	В	3.51	4.00	4.26	4.33	4.08	4.31	4.67			
L.S.D. at 5	%: A= (	0.27, B = 0.2	29, A*B=	= 0.71							
			:	Second sea	ason (2019	9)					
CCC	1500	3.73	4.40	4.73	4.40	4.47	5.07	5.30	4.59		
CCC (mg L <sup>-1</sup> )	3000	4.57	5.50	5.73	6.10	5.53	5.77	6.13	5.62		
(IIIg L )	6000	1.63	2.60	3.23	3.10	2.73	2.60	2.87	2.68		
PPP	25	4.13	5.07	5.43	5.73	4.67	4.97	5.70	5.10		
$(\text{mg L}^{-1})$	50	4.27	4.83	5.17	5.63	4.93	6.47	6.13	5.35		
(IIIg L )	75	2.93	3.17	3.27	3.70	3.07	3.50	3.90	3.36		
Mean	В	3.54	4.26	4.59	5.78	4.23	4.73	5.01			
L.S.D. at 5	%: A= (	0.30, B = 0.3	32, A*B=	= 0.79							

**Table 11.** Effect of cycocel (CCC) and paclobutrazol (PPP) in interaction with Kelpak and Milagrow on flowering period of chrysanthemum

Keipak and whiagrow on nowering period of emysanthemum											
Flowering period (days) First season (2018)											
T	1 .	Control	Kelpak (ml L <sup>-1</sup> )				grow (mg	L-1)	M A		
Treatm	ents	0	3	4	5	60	80	100	- Mean A		
CCC	1500	43.20	45.23	47.10	47.20	43.07	45.27	49.67	45.82		
$(\text{mg L}^{-1})$	3000	44.20	45.23	46.93	50.50	44.43	47.90	52.20	47.34		
(IIIg L )	6000	30.47	37.17	37.10	40.43	32.23	35.93	38.30	35.95		
PPP	25	41.13	44.53	47.00	48.43	41.90	46.43	48.83	45.47		
$(\text{mg L}^{-1})$	50	42.00	45.10	47.10	48.63	42.47	47.10	50.00	46.06		
(IIIg L )	75	24.60	30.30	33.00	36.87	34.97	38.23	40.60	34.08		
Mean		37.60	41.26	43.04	45.34	39.84	43.48	46.60			
L.S.D. at 5	5%: A=	1.38, B=1.	49, A*B=	= 3.64							
-			,	Second sea	ason (2019	9)					
CCC	1500	40.03	41.63	44.27	46.20	40.83	42.93	48.10	43.43		
$(\text{mg L}^{-1})$	3000	42.23	41.07	41.67	51.93	43.27	44.83	51.27	45.18		
(mg L )	6000	26.66	32.83	35.20	37.67	30.23	32.50	35.33	32.92		
PPP	25	39.90	40.60	42.90	45.20	40.00	43.70	46.27	42.65		
$(\text{mg L}^{-1})$	50	40.37	43.03	44.60	50.16	45.40	48.30	51.26	46.16		
(mg L )	75	26.67	29.10	35.33	38.66	37.00	39.93	40.17	35.27		
Mean	В	35.98	38.04	40.66	44.97	39.46	42.03	45.40			
L.S.D. at 5	5%: A=	1.52, B=1.	.64, A*B=	= 4.01							



**Figure 2.** Effect of different concentrations of cycocel (CCC) and paclobutrazol (PPP) on flowering aspects: flowering start (days) (A), flower number per plant (B), flower diameter (cm) (C), and flowering period (days) (D) of *D. grandiflorum* cv. 'Pink Zambla' during the seasons of 2018 and 2019. Bars represent ±Standard Deviation (n=3). Columns annotated with the same letters are not statistically different (*P*<0.05)

# Chlorophyll content (mg g<sup>-1</sup> FW)

The content of chlorophyll was increased in CCC and PPP single sprays compared to untreated plants (Figure 3A). The total chlorophyll content gradually increased with increasing concentrations of both dwarfing agents. The highest significant value of total chlorophyll (1.39 and 1.50 mg g<sup>-1</sup> FW) was recorded for the treatment of 75 mg L<sup>-1</sup> PPP while the lowest value was recorded for control plants (0.70 and 0.76 mg g<sup>-1</sup> FW) in 1<sup>st</sup> and 2<sup>nd</sup> seasons, respectively.

Respecting the interaction treatments, the total chlorophyll accumulation was enhanced due to the combined impact of growth retardants and biostimulants (Table 12). Combination of 75 mg L<sup>-1</sup> PPP with 4 or 5 ml L<sup>-1</sup> Kelpak and 80 or 100 mg L<sup>-1</sup> Milagrow recorded the highest significant value of total chlorophyll (1.52, 1.62, 1.65, and 1.49 mg g<sup>-1</sup> FW, respectively) in the first season. The medium dose of PPP (50 mg L<sup>-1</sup>) came in the second order. The

same findings were recorded in the second season. Raising the doses of Kelpak and Milagrow caused a significant increase in the accumulation of total chlorophyll.

**Table 12.** Effect of cycocel (CCC) and paclobutrazol (PPP) in interaction with Kelpak and Milagrow on total chlorophyll content in chrysanthemum

Total chlorophyll content (mg g <sup>-1</sup> FW)											
First season (2018)											
Twootma	anta.	Control	K	Celpak (ml	$L^{-1}$	Mila	grow (mg	(L-1)	Maan A		
Treatme	ents	0	3	4	5	60	80	100	Mean A		
CCC	1500	0.94	1.28	1.30	1.43	1.03	1.23	1.35	1.22		
(mg L <sup>-1</sup> )	3000	1.09	1.27	1.34	1.42	1.13	1.26	1.30	1.26		
(IIIg L )	6000	1.18	1.17	1.28	1.35	1.21	1.28	1.35	1.26		
PPP	25	0.91	1.27	1.39	1.43	1.08	1.23	1.29	1.23		
	50	1.11	1.30	1.45	1.43	1.26	1.39	1.45	1.34		
$(\text{mg L}^{-1})$	75	1.39	1.44	1.52	1.62	1.42	1.65	1.49	1.51		
Mean B		1.10	1.29	1.38	1.45	1.19	1.34	1.37			
L.S.D. at 5	%: A = 0	.077, B = 0	.084, A*	B = 0.205							
			Ş	Second sea	ason (2019	9)					
CCC	1500	1.06	1.27	1.40	1.53	1.11	1.19	1.51	1.30		
$(\text{mg L}^{-1})$	3000	1.21	1.45	1.45	1.57	1.29	1.30	1.34	1.37		
(IIIg L )	6000	1.13	1.22	1.26	1.38	1.28	1.32	1.35	1.28		
PPP	25	1.02	1.33	1.44	1.51	1.13	1.31	1.30	1.29		
	50	1.23	1.35	1.46	1.41	1.24	1.42	1.40	1.36		
$(\text{mg L}^{-1})$	75	1.50	1.56	1.66	1.80	1.42	1.67	1.57	1.60		
Mean	В	1.19	1.36	1.44	1.53	1.25	1.37	1.41			
L.S.D. at 5	%: A = 0	0.067, B = 0	.072, A*	B = 0.177							

## Carotenoids content (mg g<sup>-1</sup> FW)

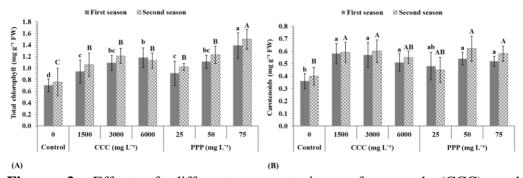
As shown in Figure (3B), low and moderate rates of CCC significantly increased the content of carotenoids. The carotenoids content reached the highest value in plants dwarfed with CCC at concentrations of 1500 and 3000 mg L<sup>-1</sup> (0.58 and 0.57 mg g<sup>-1</sup> FW, respectively) in the first season as compared to the control (0.36 mg g<sup>-1</sup> FW). In the 2<sup>nd</sup> season, the highest content of carotenoids was measured in plants treated with 1500 and 3000 mg L<sup>-1</sup> CCC, 50 and 75 mg L<sup>-1</sup> PPP (0.59, 0.60, 0.62, and 0.58 mg g<sup>-1</sup> FW, respectively) compared to control (0.40 mg g<sup>-1</sup> FW).

Data displayed in Table (13) show that combination treatments did not significantly enhance the accumulation of carotenoids in chrysanthemum leaves when compared with the single application of the dwarfing agents except for 25 mg L<sup>-1</sup> PPP. A significant increase in the carotenoids content was observed when Kelpak was sprayed on plants treated with 25 mg L<sup>-1</sup> PPP. The results revealed that Kelpak was more effective than Milagrow in increasing the

carotenoids content in the leaves of plants treated with CCC or PPP. The highest content of carotenoids (0.66 and 0.70 mg g<sup>-1</sup> FW) was measured for the combination of 1500 mg L<sup>-1</sup> CCC with 5 ml L<sup>-1</sup> Kelpak in the 1<sup>st</sup> and 2<sup>nd</sup> seasons, respectively.

**Table 13.** Effect of cycocel (CCC) and paclobutrazol (PPP) in interaction with Kelpak and Milagrow on carotenoids content in chrysanthemum

Carotenoids content (mg g <sup>-1</sup> FW)									
First season (2018)									
Treatments		Control	Kelpak (ml L <sup>-1</sup> )			Milagrow (mg L <sup>-1</sup> )			- Mean A
		0	3	4	5	60	80	100	- Mean A
CCC (mg L <sup>-1</sup> )	1500	0.58	0.63	0.62	0.66	0.44	0.43	0.45	0.54
	3000	0.57	0.58	0.63	0.59	0.45	0.46	0.48	0.53
	6000	0.51	0.52	0.45	0.43	0.39	0.40	0.40	0.44
PPP (mg L <sup>-1</sup> )	25	0.48	0.52	0.53	0.55	0.42	0.47	0.49	0.49
	50	0.54	0.53	0.57	0.56	0.43	0.49	0.50	0.51
	75	0.52	0.53	0.53	0.56	0.47	0.51	0.50	0.52
Mean B		0.53	0.55	0.55	0.56	0.43	0.46	0.47	
L.S.D. at 5%: A= 0.053, B= 0.057, A*B= 0.139									
Second season (2019)									
CCC (mg L <sup>-1</sup> )	1500	0.59	0.61	0.65	0.70	0.45	0.46	0.44	0.56
	3000	0.60	0.60	0.65	0.63	0.53	0.51	0.55	0.58
	6000	0.55	0.57	0.54	0.48	0.44	0.47	0.45	0.50
PPP (mg L <sup>-1</sup> )	25	0.45	0.54	0.57	0.61	0.45	0.50	0.53	0.52
	50	0.62	0.52	0.55	0.58	0.50	0.54	0.56	0.55
	75	0.58	0.57	0.56	0.58	0.45	0.53	0.49	0.54
Mean B		0.57	0.57	0.59	0.60	0.47	0.50	0.50	
L.S.D. at 5%: A= 0.046, B= 0.050, A*B= 0.122									



**Figure 3.** Effect of different concentrations of cycocel (CCC) and paclobutrazol (PPP) on total chlorophyll (A), and carotenoids (B) content (mg g<sup>-1</sup> FW) of *D. grandiflorum* cv. 'Pink Zambla' during the seasons of 2018 and 2019. Bars represent  $\pm$ Standard Deviation (n=3). Columns annotated with the same letters are not statistically different (P<0.05)

#### **Discussion**

To produce chrysanthemum plants as potted plant, improving the vegetative growth characteristics such as dwarfing height and increasing branching is one of the most desirable traits to obtain a dwarfed plant full of flowering heads. Application of PGRs (CCC and PPP) on D. grandiflorum cv. 'Pink Zambla' caused a significant inhibition in the stem height compared to untreated plants. The commercially desirable stem length (around 30 cm) can be achieved by using moderate rates of CCC (3000 mg L<sup>-1</sup>) and PPP (50 mg L<sup>-1</sup>) 1). The mechanism of the reduction in plant height here appears to be related to the effect of PGRs in slowing cell division and restricting cell elongation (Magnitskiy et al., 2006; Karunananda and Peiris, 2010). CCC is known to completely inhibit the biosynthesis of gibberellins (GA) (Li et al., 2023); the main plant hormone responsible for cell elongation. To achieve a desirable plant height of high quality suitable for commercial production of potted chrysanthemum, combinations of PGRs and growth stimulants; SE (Kelpak) and BL (Milagrow), were applied. The interaction of CCC or PPP with Kelpak or Milagrow has enhanced these traits. In this connection, Ghoneim (2016), found that *Pelargonium zonale* plants treated with foliar spray of Milagrow at 80 mg L<sup>-1</sup> and Kelpak at 4 ml L<sup>-1</sup> recorded the highest plants. Moreover, Zong et al. (2019) reported that application of BRs increased plant height of Leymus chinensis due to improvements in physiochemical. The most appropriate result here (25-35 cm for stem length) is obtained due to applying moderate doses of the dwarfing agents in combination with any rate of both stimulants.

Decreasing leaves number in the dwarfed plants may be attributed to the resulting inhibition of stem height under the influence of PGRs. However, leaf number was increased when the stunted plants were sprayed with biostimulants. This increment may be related to the positive impact of biostimulants on increasing stem length and branching, as well as providing the nutritional growth necessities which were contained in these extracts. Moreover, it has been proven that these bio-stimulants act as anti-stress (Samira *et al.*, 2012; Behnamnia, 2015). Promotion of shoot growth and leaf number is an indicator that BL (Milagrow) mitigated the negative impact of the dwarfing agents on the potted chrysanthemum.

PGRs are known to activate lateral buds to grow and fill in with a greater number of branches (Benjawan *et al.*, 2007). In the current experiment, low and moderate levels of PGRs improved branching. Abbas *et al.* (2007) found that CCC levels (500, 1000 and 1500 mg L<sup>-1</sup>) promoted shoot development in *Rosa damascene* and the highest branching was recorded for 1500 mg L<sup>-1</sup>CCC. Also, the maximum branching in *Tagetes erecta* was reported at 2000 mg L<sup>-1</sup> of CCC

(Khan et al., 2012). The enhancement of branching due to the application of low and medium levels of CCC and PPP may be attributed to the inhibitory action of these growth regulators on cell division in the apical bud, which may arrest the growth of the main axis and enhance lateral production and may also be due to the inhibition of auxin activity in the apical bud as it acts as an auxin antagonist (Prashanth et al., 2006; Di Benedetto and Molinari, 2007). Hence, special care is needed to establish bushy and dwarfed chrysanthemum plants to suit market specifications mainly in terms of plant height and branche number. The lowest significant number of branches was recorded with the single application of high levels of CCC (6000 mg L<sup>-1</sup>) and PPP (75 mg L<sup>-1</sup>), due to the toxic effect on the plant cell. However, the application of the growth biostimulants mitigated this toxicity and increased branching. Mohamed (2020) showed that BL at 50 mg L<sup>-1</sup> and its combination with mycorrhizal fungi increased the number of branches in fennel, however, SE (2 ml L<sup>-1</sup>), as well as its combination with mycorrhizal fungi resulted in the highest values for branch count. The effect of BL (Milagrow®) on vegetative parameters may be due to the improvement of cell growth, differentiation, division and enlargement, alteration of membrane potential, and metabolism of nucleic acids and proteins (M üssig, 2005; Dehghan *et al.*, 2020).

As expected, the biomass FW and DW of chrysanthemum plants treated with a high rate of PGRs alone were decreased because of the severe reduction in plant height, branche number, and leaf number compared to the non-sprayed plants. Al-Shaer (2004) reported a similar observation that CCC increased the number of branches per plant but reduced both FW and DW in Grindelia camporum. Enhancing the FW with 25 and 50 mg L<sup>-1</sup> PPP implies that the plants may have absorbed more water and nutrients. The decrease in DW was linearly related to the increase in the concentration of CCC and PPP. North et al. (2010) on Dombeya burgessiae reported the same observation where the FW and DW of plants were severely reduced with the increase of CCC concentration. Spraying plants with the bio-stimulants increased the FW and DW of the stunted plants. In general, the FW and DW were increased by increasing the concentrations of both bio-stimulants. This observation is in harmony with that recorded by Ferreira dos Santos et al. (2019) on ornamental sunflower. This could be related to the accumulation of phytonutrients and water in plant tissues. Similarly, Zong et al. (2019) found that the enhancement in physiochemical attributes due to BR application increased FW and DW of Leymus chinensis. BL is well known to protect plants exposed to abiotic stress and enhance plant growth and biomass FW and DW (Fariduddin et al., 2014).

The stimulating impact of SE on vegetative growth characteristics may be attributed to its primary action in promoting cell division and elongation as it

contains high amounts of natural nutrients such as N, P, K, Ca, Mg, S, Fe, Cu, Mn, B and Mo, natural hormones (cytokinins, auxins, and gibberellins), amino acids, and vitamins (Begum *et al.*, 2018). Moreover, studies with *Arabidopsis thaliana* cells revealed the ability of BRs to stimulate cell expansion which was accompanied by the amplitude of the hyperpolarization of the cell membrane (Zhang *et al.*, 2005).

Flowering characteristics, such as the beginning of flowering, flower number per plant, flower diameter, and flowering period are important characteristics that producers of potted plants strive to enhance and increase. PGRs applied separately to chrysanthemum caused significantly earlier flowering compared to the control. Close to our results, Vaghasia and Polara (2016) found that CCC at 2500 mg  $L^{-1}$  gave the earliest flowering date in C. morifolium cv. IIHR-6. In contrast, Ghatas (2016) found that treatments of CCC (1000, 2000, and 3000 mg L<sup>-1</sup>) and PPP (20, 40, and 60 mg L<sup>-1</sup>) delayed the flowering of C. frutescence plants as compared with untreated plants, which indicates that the effect of these PGRs depends on many factors, including the plant species. Early flowering is a desirable trait for the producer to reduce production costs. The number of days required for flowering has gradually decreased with increasing concentration of the bio-stimulants sprayed on the dwarfed plants. In parallel with our results, Kandil et al. (2007) found that foliar application of 50 mg L<sup>-1</sup> GA3 + 15 mg L<sup>-1</sup> BL + 40 mg L<sup>-1</sup> Kin promoted flowering start in Rosa hybrida. The presence of very high levels of BRs in pollen and seeds indicates the involvement of BRs in the regulation of reproductive growth and reproduction (Ali, 2017). Müssig (2005) suggested that BRs can determine branching and flower formation by modulating metabolic pathways and nutrient allocation or interacting with other signaling pathways. The early flowering of chrysanthemum plants sprayed with Kelpak may be caused by the early development of the plant due to the availability of phytonutrients.

As for the commercial production of potted plants, producing a plant full of flowers as much as possible is one of the most important goals that pot plant producers seek. Plants treated with the PGRs at moderate rates produced a higher number of flowers. Also, Vaghasia and Polara (2016) found that 2500 mg L<sup>-1</sup> of CCC enhanced flower number in *C. morifolium* cv. IIHR-6. Moreover, Ghatas (2016) found that the highest flower number was recorded for *C. frutescence* plants subjected to 60 mg L<sup>-1</sup> of PPP. A high concentration of dwarfing factors led to a sharp decrease in flower number, which may be due to the decrease in the number of shoots for these treatments, as previously mentioned. The number of flowers of chrysanthemum received both biostimulants and dwarfing compounds was better than that of plants treated with

PGRs separately. This increase in the number of flowers is attributed to the increase in branche number. Kandil *et al.* (2007) noticed an increase in the flower number in *Rosa hybrida* when a conjunction of 50 mg L<sup>-1</sup> GA3 + 15 mg L<sup>-1</sup> BL + 40 mg L<sup>-1</sup> Kin was applied. Moreover, a stimulating effect of seaweed liquid fertilizer on flower number in *Tagetes erecta* was also reported by Sridhar and Rengasamy (2010).

Application of CCC and PPP each alone reduced the flower diameter. In conformity with this observation, Mishra and Yadava (2011) recorded a decrease in flower size and flower stalk length by increasing PPP concentration in China aster cv. Poornima. This may be related to the decrease in leaf number and therefore fewer overall nutrients needed to increase the flower size. However, a slight increment in flower size was recorded for moderate application of CCC, which may be due to the increased content of chlorophyll and the rate of photosynthesis, and thus the ultimately higher manufacture of photosynthetic increased flower size as reported by Singh et al. (2018). Similar results were recorded on *Primula forbesii* plants dwarfed with CCC (Zhang et al., 2020). The flower size was significantly improved when the growth stimulants and dwarfing agents were crossed. The positive effect of BL (Milagrow) on flower diameter could be due to the promoting of petal growth through cell expansion. In this concept area, Huang et al. (2017) found that BL promoted petal growth in Gerbera hybrida by elongating cells in the central and basal regions of the petals. The authors demonstrated that such effect on petal growth was greater than that of GA. Moreover, increasing flower diameter with Kelpak and Milagrow application could be correlated with the increase in photosynthesis rate (Fariduddin et al., 2014).

Only 3000 mg L<sup>-1</sup> of CCC significantly enhanced the flowering period in the second season. In line with these results, Kumar *et al.* (2019) found that CCC at 2500 mg L<sup>-1</sup> recorded the early flowering and maximum duration of flowering in *Nerium odorum* L. On chrysanthemum cv. Birbal Sahni, Singh *et al.* (2018) found that the maximum flowers yield, and shelf life of flowers were achieved with 5000 mg L<sup>-1</sup> of CCC. However, in our study, high concentration of CCC (6000 mg L<sup>-1</sup>) decreased the flowering period. This may be due to the difference in the cultivar and agricultural conditions. The flowering period was positively prolonged for the combination application. Bosila *et al.* (2016) on *Pelargonium zonale* 'Serena' reported that application of Kelpak gave the highest value of flowering period compared with chemical fertilization and control treatments.

The enhancement of flowering traits as a result of spraying SE products could be due to promoting plant growth (Mohamed, 2020). Moreover, the use of SE led to the activity of enzymes and thus increased biological processes

within the plant cell, resulting in increased plant growth and productivity (EL Boukhari *et al.*, 2020). Recently, Sheng *et al.* (2022) reported that BL signaling acts as a growth enhancer during vegetative and reproductive development in lotus (*Nelumbo nucifera*). The authors observed growth-promoting phenotypes and long-lasting flowering because of the exogenous application of 28-epihomobrassinolide.

It is known that biosynthesis and accumulation of pigments in plant tissues are affected by many factors, including nutrients and various stress factors, such as chemical compounds used in dwarfing potted plants (Abdel-Gawad, 2016; Ghoneim, 2016; Abbas, 2017). The content of chlorophyll was increased in chrysanthemum plants treated with PGRs which has also been confirmed in C. frutescence (Ghatas, 2016) and Tabernaemontana coronaria (Youssef and Abd El-Aal, 2013). This may be due to the inhibitory effect of PGRs which produced smaller cells and subsequently increased the concentration of chlorophyll within the reduced cell volume (Thakur et al., 2006). In addition, Tsegaw et al. (2005) suggested that the increased accumulation of chlorophyll in potato leaves upon application of PPP was due to enhanced chlorophyll biosynthesis as well as to more densely spaced chloroplasts per unit leaf area. It was reported that treatment with CCC and PPP resulted in an increase in chlorophyll content and photosynthesis rate, and consequently, increased the accumulation of carbohydrates in plant leaves (Zheng et al., 2012). Thus, this may be the reason for the increase in plant biomass, flower number, and the prolongation of the flowering period in some treatments in the current investigation compared to the control. Kelpak acted as a stronger growth stimulant than Milagrow in increasing the chlorophyll and carotenoids content in the stunted plants. A similar observation was recorded by Mohamed (2020) on the Dutch fennel as SE (2 ml L<sup>-1</sup>) and its combination with mycorrhizal fungi recorded the highest values of chlorophylls compared to other stimulants i.e., amino acids, BL, humic acid, and salicylic acid. Carotenoids are a class of terpenoids with multiple functions such as photosynthetic pigments and plant protection under different stresses (Salam et al., 2023). Hu et al. (2013) observed that the photosynthetic capacity, net photosynthetic rate, light saturation rate, and quantitative efficiency of PSII were improved in Leymus chinensis treated with a mixture of abscisic acid and BL. Furthermore, under abiotic stress, BL can stimulate chlorophyll synthesis (Yadava et al., 2016). Increased level of bio-stimulants was accompanied by an increase in chlorophyll content. In this regard, Al-Dulaimy et al. (2021) found that Gazania splendens treated with SE at 4 ml L<sup>-1</sup> recorded the highest leaf content of chlorophyll and total carbohydrates compared to control and low level. The stimulatory effects of the PGRs on enhancing the biosynthesis of carotenoids have been confirmed by Lodeta *et al.* (2010) and Abbas (2017) on poinsettia plants, and Abdel-Moniem (2016) on sunflower plant.

In conclusion, when producing potted plants, controlling stem height for proper length, good branching and a consistent vegetative shape with the flowers is often necessary to achieve the desired size and shape for marketing ornamental crops. In the current study, moderate and low concentrations of growth retardants (CCC and PPP) had the desired effects in manipulating the growth parameters of D. grandiflorum cv. 'Pink Zambla', and thus shaped as a potted plant. Concerning the key characteristics of potted chrysanthemum, the best results were recorded in plants treated by a combination of 3000 mg L<sup>-1</sup> CCC or 50 mg L<sup>-1</sup> PPP with different rates of Kelpak or Milagrow. This makes us recommend the use of growth stimulants along with growth inhibitors when the purpose is to produce potted dwarfed chrysanthemum plants with high flowering quality that increase their commercial value and meet the needs of the consumer and market of potted flowering plants. Thus, combining growth stimulants with dwarfing agents may be a good option to mitigate cytotoxicity caused by growth retardants. On the other hand, the farmers' improper use of inorganic fertilizers to improve plant growth and productivity is harmful to the environment and human health. The use of bio-stimulants such as SE (Kelpak) and brassinosteroid (Milagrow) in agriculture could be an efficient and ecofriendly alternative to chemical fertilizers.

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#### References

- Abbas, M. M., Ahmad, S. and Anwar, R. (2007). Effect of growth retardants to break apical dominance in *Rosa damascena*. Pakistan Journal of Agricultural Sciences, 44:524-529.
- Abbas, M. N. (2017). Effect of some cultural treatments on the vegetative growth and flowering in Euphorbia pulcherrima plant. M.Sc. Thesis. Faculty of Agriculture, Al-Azhar University, Cairo, Egypt.
- Abdel-Gawad, A. I. M. (2016). Some factors affecting the growth and flowering of Chrysanthemum indicum plant. M.Sc., Thesis, Faculty of Agriculture, Al-Azhar University, Cairo, Egypt.
- Abdel-Moniem, A. M. (2016). Effect of some growth retardants on growth and flowering of *Helianthus annuus* L. cv. Sunrich Orange Summer 981V plants. B- Application of ancymidol, daminozide and ethephon in combinations. Scientific Journal of Flowers and Ornamental Plants, 3:119-134.
- Abou Elhassan, M. H., Bosila, H. A., Hamza, M. A., Elateeq, A. A. and Abdel-Gawad, A. I. (2021). Effect of cycocel and paclobutrazol on the dwarfing characteristics of *Chrysanthemum indicum* L. Al-Azhar Journal of Agricultural Research, 46:41-50.

- Al-Dulaimy, A. F. Z., Alani, M. H. I., Awad, I. M., Khudair, R. I. A., Salim, H. A. and Jasem, S. M. (2021). Response of Gazania plants (*Gazania splendens* L.) to seaweed extract (Tecamin Algae) and nano stimulator (Proteck CalBor). IOP Conference Series: Earth and Environmental Science. IOP Publishing, 910:012052.
- Al-Shaer, A. E. I. (2004). Effect of some hormonal treatments and cultural medium on growth, flowering and resin content of Grindelia camporum green plants. Thesis, Faculty of Agriculture, Al-Azhar University, Cairo, Egypt.
- Ali, B. (2017). Practical applications of brassinosteroids in horticulture—some field perspectives. Scientia Horticulturae, 225:15-21.
- Anton, I., Dorneanu, A., Dana, D. and Coteţ, V. (2010). Agriculture development of Romania in EU integration conditions by using unconventional means of fertilization. Research Journal of Agricultural Science, 42:8-13.
- Asrar, A.-W., Elhindi, K. and Abdel-Salam, E. (2014). Growth and flowering response of chrysanthemum cultivars to Alar and slow-release fertilizer in an outdoor environment. Journal of Food, Agriculture & Environment, 12:963-971.
- Begum, M., Bordoloi, B. C., Singha, D. D. and Ojha, N. J. (2018). Role of seaweed extract on growth, yield and quality of some agricultural crops: A review. Agricultural Reviews, 39:321-326.
- Behnamnia, M. (2015). Protective roles of brassinolide on tomato seedlings under drought stress. International Journal of Agriculture and Crop Sciences, 8:455-462.
- Benjawan, C., Chutichudet, P. and Chanaboon, T. (2007). Effect of chemical paclobutrazol on growth, yield and quality of okra (*Abelmoschus esculentus* L.) Har lium cultivar in northeast Thailand. Pakistan Journal of Biological Sciences, 10:433-438.
- Bosila, H. A., Zeawail, M. E.-F., Sleem, A. H., Hamza, M. A. and Ghoneim, A. H. (2016). Influence of seaweed extract (Kelpak) on flowering in *Pelargonium zonale* L. Var 'Serena' plant. Journal of Biological Chemistry and Environmental Sciences, 11:529-538
- Dehghan, M., Balouchi, H., Yadavi, A. and Zare, E. (2020). Improve wheat (*Triticum aestivum*) performance by brassinolide application under different irrigation regimes. South African Journal of Botany, 130:259-267.
- Di Benedetto, A. and Molinari, J. (2007). Influence of river waste-based media on efficacy of paclobutrazol in inhibiting growth of *Petunia* x *hybrida*. International Journal of Agricultural Research, 2:289-295.
- Eid, F. S., El-Kholy, M. F. and Hosny, S. S. (2016). Effect of foliar sprays application of Milagrow on yield and fruit quality of avocado tree cv. "Fuerte". Journal of Plant Production, Mansoura University, 7:1495-1499.
- EL Boukhari, M. E., Barakate, M., Bouhia, Y. and Lyamlouli, K. (2020). Trends in seaweed extract based biostimulants: Manufacturing process and beneficial effect on soil-plant systems. Plants, 9:359.
- Fariduddin, Q., Yusuf, M., Begum, M. and Ahmad, A. (2014). 28-homobrassinolide protects photosynthetic machinery in Indian mustard under high temperature stress. Journal of Stress Physiology & Biochemistry, 10:181-194.
- Ferreira dos Santos, P. L., Zabotto, A. R., Jordão, H. W. C., Boas, R. L. V., Broetto, F. and Tavares, A. R. (2019). Use of seaweed-based biostimulant (*Ascophyllum nodosum*) on ornamental sunflower seed germination and seedling growth. Ornamental Horticulture, 25:231-237.
- Ghatas, Y., Ali, M., Elsadek, M. and Mohamed, Y. (2021). Enhancing growth, productivity and artemisinin content of *Artemisia annua* L. plant using seaweed extract and micronutrients. Industrial Crops and Products, 161:113202.

- Ghatas, Y. A. A. (2016). Influence of paclobutrazol and cycocel sprays on the growth, flowering and chemical composition of potted *Chrysanthemum frutescens* plant. Annals of Agricultural Science, Moshtohor, 54:355-364.
- Ghoneim, A. H. M. (2016). Some factors affecting the growth and flowering of Pelargonium zonale plant. (Master Thesis), Faculty of Agriculture, Al-Azhar University, Cairo, Egypt.
- Hu, Y.-J., Shi, L.-X., Sun, W. and Guo, J.-X. (2013). Effects of abscisic acid and brassinolide on photosynthetic characteristics of *Leymus chinensis* from Songnen Plain grassland in Northeast China. Botanicals Studies, 54:42.
- Huang, G., Han, M., Yao, W. and Wang, Y. (2017). Transcriptome analysis reveals the regulation of brassinosteroids on petal growth in *Gerbera hybrida*. PeerJ, 5:e3382.
- Kandil, M. M., Shalaby, M. A. and Mahgoub, M. H. (2007). Effect of some growth regulators on the levels of endogenous hormones chemical and constituents of rose plant. American-Eurasian Journal of Agricultural and Environmental Sciences, 2:720-730.
- Kanwar, M. K., Bakshi, P., Sharma, P., Kour, J., Singh, A. D., Dhiman, S., Ibhrahim, M., Mir, B. A., Ahammed, G. J., Zhou, J. and Bhardwaj, R. (2022). *Brassinosteroids in plant reproductive development*. In: Ahammed, G. J., Sharma, A., & Yu, J. (Eds) Brassinosteroids in Plant Developmental Biology and Stress Tolerance. Academic Press, pp. 105-130.
- Karunananda, D. P. and Peiris, S. E. (2010). Effects of pinching, cycocel and B-nine treatments on branching habit of pot poinsettia (*Euphobia pulcherrima* Willd). Tropical Agricultural Research, 21:284-292.
- Khan, M. I., Muzamil, S., Abid, M., Hassan, A. and Mathew, B. (2012). Effect of different levels of cycocel and maleic hydrazide on growth and flowering of African Marigold (*Tagetes erecta* L.) cv. PUSA NARANGI GAINDA. The Asian Journal of Horticulture, 7:294-296.
- Kumar, S., Haripriya, K., Kumar, S., Muraleedharan, A. and Kamalakannan, S. (2019). Effect of cycocel on growth, flowering and yield of nerium (*Nerium odorum* L.). Journal of Pharmacognosy and Phytochemistry, 8:2226-2228.
- Li, Q., Zhang, L., Cao, S., Li, J. A., Yan, J., Xiong, L., Wang, F. and He, J. (2023). Dwarfing effect of plant growth retarders on *Melaleuca alternifolia*. Forests, 14:732.
- Lichtenthaler, H. K. (1987). Chlorophylls and carotenoids: pigments of photosynthetic biomembranes. In: Methods in enzymology. Elsevier, Academic Press, 148:350-382.
- Lodeta, K. B., Ban, S. G., Perica, S., Dumičić, G. and Bućan, L. (2010). Response of poinsettia to drench application of growth regulators. Journal of Food, Agriculture & Environment, 8:297-301.
- Mabberley, D. J. (2008). Mabberley's Plant-book: a portable dictionary of plants, their classifications and uses. Cambridge University Press.
- Magnitskiy, S. V., Pasian, C. C., Bennett, M. A. and Metzger, J. D. (2006). Controlling plug height of *Verbena*, *Celosia*, and *Pansy* by treating seeds with paclobutrazol. Hortscience, 41:158-161.
- Mishra, D. K. and Yadava, L. P. (2011). Influence of paclobutrazol application on the flower size and yield of China aster (*Callistephus chinensis* (L.) Nees). Journal of Applied Horticulture, 13:147-149.
- Mohamed, Y. F. Y. (2020). Impact of some growth stimulants in cooperation with arbuscular mycorrhizal fungi on growth, productivity and chemical constituents of Dutch fennel plant. Scientific Journal of Flowers and Ornamental Plants, 7:303-319.
- Müssig, C. (2005). Brassinosteroid-promoted growth. Plant Biology, 7(2):110-117.

- Nada, R. S., Ashmawi, A. E., Mady, E., Randhir, T. and Elateeq, A. A. (2022). Effect of organic manure and plant growth promoting microbes on yield, quality and essential oil constituents of fennel bulb (*Foeniculum vulgare* Mill.). Journal of Ecological Engineering, 23:149-164.
- North, J. J., Laubscher, C. P. and Ndakidemi, P. A. (2010). Effect of the growth retardant Cycocel® in controlling the growth of *Dombeya burgessiae*. African Journal of Biotechnology, 9:4529-4533.
- Oyoo, J., Nyongesa, M., Mbiyu, M. and Lungaho, C. (2010). Organic farming: effect of Kelpak and Earthlee on the yield of Irish potatoes. Proceedings of the 12th Kari Biennial Scientific Conference "Transforming Agriculture for improved livelihoods through Agricultural Product Value Chains, pp 8-12.
- Pobudkiewicz, A. (2014). Effect of growth retardant on some morphological and physiological traits of chrysanthemum. Polish Journal of Natural Sciences, 29:291-306.
- Prashanth, P., Reddy, S. A. and Srihari, D. (2006). Studies on the effect of certain plant growth regulators on growth of Floribunda roses (*Rosa hybrida* L.). Orissa Journal of Horticulture, 34:78-82.
- Salam, U., Ullah, S., Tang, Z.-H., Elateeq, A. A., Khan, Y., Khan, J., Khan, A. and Ali, S. (2023). Plant metabolomics: An overview of the role of primary and secondary metabolites against different environmental stress factors. Life, 13:706.
- Samira, I. M.-H., Dridi-Mouhandes, B., Mansour-Gueddes, S. and Denden, M. (2012). 24-Epibrassinolide ameliorates the adverse effect of salt stress (NaCl) on pepper (*Capsicum annuum* L.). Journal of Stress Physiology & Biochemistry, 8:232-240.
- Seadh, S. E., Attia, A. N. E., Badawi, M. A. and El-Hety, M. S. E. (2012). Response of seed yield and its components of safflower to sowing dates, nitrogen fertilizer levels and times of foliar application with milagrow. Journal of Biological Sciences, 12:342-348.
- Sheng, J., Li, X. and Zhang, D. (2022). Gibberellins, brassinolide, and ethylene signaling were involved in flower differentiation and development in *Nelumbo nucifera*. Horticultural Plant Journal, 8:243-250.
- Singh, J., Nigam, R., Singh, R., Kumar, A. and Kumar, A. (2018). Effect of gibberellic acid and cycocel on growth, flowering and yield of chrysanthemum (*Dendranthema grandiflora* Ramat) cv. Birbal Sahni. Journal of Pharmacognosy and Phytochemistry, SP1:2753-2758.
- Snedecor, G. W. and Cochran, W. G. (1980). Statistical Methods. 7th ed. Iowa State University Press, Ames, Iowa.
- Soliman, M. N., Toaima, N. M. and Mahmoud, S. M. (2022). Effect of different levels of cycocel on the vegetative growth and flowering of *Gardenia jasminoides* J. Ellis plant. Al-Azhar Journal of Agricultural Research, 47:46-56.
- Sridhar, S. and Rengasamy, R. (2010). Effect of seaweed liquid fertilizer on the growth, biochemical constituents and yield of *Tagetes erecta*, under field trial. Journal of Phytology, 2:61-68.
- Thakur, R., Sood, A., Nagar, P. K., Pandey, S., Sobti, R. C. and Ahuja, P. S. (2006). Regulation of growth of *Lilium* plantlets in liquid medium by application of paclobutrazol or ancymidol, for its amenability in a bioreactor system: growth parameters. Plant Cell Reports, 25:382-391.
- Toaima, N., Mahmoud, S., Hamza, M. and Nour el deen, M. A. (2018). Influence of different levels of Ethephon on the growth and flowering of *Euphorbia pulcherrima* var. 'Freedom Red'. Arabian Journal of Medicinal & Aromatic Plants, 4:45-55.
- Tsegaw, T., Hammes, S. and Robbertse, J. (2005). Paclobutrazol-induced leaf, stem, and root anatomical modifications in potato. Hortscience, 40:1343-1346.

- Vaghasia, M. and Polara, N. D. (2016). Effect of plant growth retardants on growth, flowering and yield of chrysanthemum (*Chrysanthemum morifolium* Ramat.) cv. IIHR-6. Malaysian Journal of Medical and Biological Research, 3:99-104.
- Vijayakumar, S., Nair, S. A., Nair, A. K., Laxman, R. H. and Kalaivanan, D. (2021). Influence of phenophase based irrigation and fertigation schedule on vegetative performance of chrysanthemum (*Dendranthema grandiflora* Tzelev.) var. Marigold. Journal of Horticultural Sciences, 16:222-233.
- Wickramasinghe, N. P. C. H., Seran, T. H. and Senarathne, M. M. D. J. (2021). Flowering attributes of *Henckelia* Royal Queen influenced by pinching and Paclobutrazol application. Bangladesh Journal of Scientific and Industrial Research, 56:177-184.
- Xu, L. and Geelen, D. (2018). Developing biostimulants from agro-food and industrial by-products. Frontiers in Plant Science, 9:1567.
- Yadava, P., Kaushal, J., Gautam, A., Parmar, H. and Singh, I. (2016). Physiological and biochemical effects of 24-epibrassinolide on heat-stress adaptation in maize (*Zea mays* L.). Natural Science, 8:171-179.
- Youssef, A. S. M. and Abd El-Aal, M. M. M. (2013). Effect of paclobutrazol and cycocel on growth, flowering, chemical composition and histological features of potted *Tabernaemontana coronaria* Stapf plant. Journal of Applied Sciences Research, 9:5953-5963.
- Youssef, F. S., Eid, S. Y., Alshammari, E., Ashour, M. L., Wink, M. and El-Readi, M. Z. (2020). *Chrysanthemum indicum* and *Chrysanthemum morifolium*: chemical composition of their essential oils and their potential use as natural preservatives with antimicrobial and antioxidant activities. Foods, 9:1460.
- Zhang, M., Yang, J., Pan, H. and Pearson, B. J. (2020). Dwarfing effects of chlormequat chloride and uniconazole on potted baby primrose. HortTechnology, 30:536-543.
- Zhang, Z., Ramirez, J., Reboutier, D., Brault, M., Trouverie, J., Pennarun, A.-M., Amiar, Z., Biligui, B., Galagovsky, L. and Rona, J.-P. (2005). Brassinosteroids regulate plasma membrane anion channels in addition to proton pumps during expansion of *Arabidopsis thaliana* cells. Plant Cell Physiology, 46:1494-1504.
- Zheng, R.-r., Wu, Y. and Xia, Y.-p. (2012). Chlorocholine chloride and paclobutrazol treatments promote carbohydrate accumulation in bulbs of *Lilium* Oriental hybrids 'Sorbonne'. Journal of Zhejiang University SCIENCE B, 13:136-144.
- Zong, X., Dong, Y., Shahid, M., Ahmad-Anjum, S., Shakoor, A., Lv, J., Li, N., He, X., Xu, Y., Wu, X. and Wang, S. (2019). Improving growth and physiochemical attributes of *Leymus chinensis* through exogenous brassinolide, urea and potassium dihydrogen phosphate in arid grassland. Agrociencia, 53:99-114.

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